organic compounds

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Methyl 2-acetamido-2-deoxy-β-Dglucopyranoside dihydrate and methyl 2-formamido-2-deoxy-β-D-glucopyranoside

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Methyl 2-acetamido-2-deoxy- β -D-glucopyranoside (β -GlcN-AcOCH₃), (I), crystallizes from water as a dihydrate, $C_0H_{17}NO_6H_2O_6$ containing two independent molecules [denoted (IA) and (IB)] in the asymmetric unit, whereas the crystal structure of methyl 2-formamido-2-deoxy- β -D-glucopyranoside (β -GlcNFmOCH₃), (II), C₈H₁₅NO₆, also obtained from water, is devoid of solvent water molecules. The two molecules of (I) assume distorted ${}^{4}C_{1}$ chair conformations. Values of φ for (IA) and (IB) indicate ring distortions towards $B_{C2,C5}$ and $^{C3,O5}B$, respectively. By comparison, (II) shows considerably more ring distortion than molecules (IA) and (IB), despite the less bulky N-acyl side chain. Distortion towards $B_{C2,C5}$ was observed for (II), similar to the findings for (IA). The amide bond conformation in each of (IA), (IB) and (II) is *trans*, and the conformation about the C-N bond is *anti* (C-H is approximately anti to N-H), although the conformation about the latter bond within this group varies by $\sim 16^{\circ}$. The conformation of the exocyclic hydroxymethyl group was found to be gt in each of (IA), (IB) and (II). Comparison of the X-ray structures of (I) and (II) with those of other GlcNAc mono- and disaccharides shows that GlcNAc aldohexopyranosyl rings can be distorted over a wide range of geometries in the solid state.

Comment

Acylation is an important covalent modification that affects the biological functions of saccharides and other biomolecules. Two types of saccharide acylation are common in biological systems, *N*- and *O*-acylation, and recent work has shown that different *O*-acylation patterns affect biological function. For example, this type of covalent control has been described recently for glycopeptidolipids involved in signaling through Toll-like receptors (Sweet *et al.*, 2008). *N*-Acylation is found in biologically relevant monosaccharides, such as *N*-acetyl-D- glucosamine, N-acetyl-D-galactosamine and N-acetylneuraminic acid. It has been shown recently that cis-trans isomerization (CTI) of the amide bond in N-acylated sugars can be detected in aqueous solution, with the cis/trans ratio dependent on, among other factors, the anomeric configuration of the saccharide (Hu, Zhang et al., 2010). For example, $K_{translcis}$ is ~60 for methyl N-acetyl- α -D-glucosaminide and \sim 38 for methyl *N*-acetyl- β -D-glucosaminide at 326 K. In support of NMR studies of saccharide CTI and of the parameterization of NMR J couplings within saccharide exocyclic N-acyl fragments (Hu, Carmichael & Serianni, 2010), we undertook the crystallization of methyl 2-acetamido-2-deoxy- β -D-glucopyranoside dihydrate, (I), and methyl 2-formamido-2-deoxy- β -D-glucopyranoside, (II). Their crystal structures, reported here, are compared with the structurally related compounds N-acetyl- α -D-glucosamine (2acetamido-2-deoxy- α -D-glucopyranose), (III) (α -GlcNAcOH; Mo & Jensen, 1975), β -chitobiose [2-acetamido-2-deoxy- β -Dglucopyranosyl- $(1 \rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranose], (IV) (β -GlcNAcOH and β -GlcNAcOR; Mo, 1979), α -chitobiose [2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy-α-D-glucopyranose], (V) (α-GlcNAcOH and β -GlcNAcOR; Mo & Jensen, 1978) and methyl β -Dglucopyranoside, (VI) (β -GlcOCH₃; Jeffrey & Takagi, 1977).



Crystals of (I) and (II) were obtained from aqueous solutions by slow evaporation at room temperature. Compound (I) crystallizes with two independent molecules in the asymmetric unit [denoted (IA) and (IB)] and two solvent water molecules



Figure 1

Labeling schemes for (a) molecule (IA) with water molecules and (b) molecule (IB). Displacement ellipsoids are drawn at the 50% probability level.

(Fig. 1*a* and 1*b*), whereas the unit cell of (II) contains only one molecule of the saccharide and no solvent water molecules (Fig. 2).

A comparison of selected structural parameters in compounds (I)–(VI) is shown in Table 1. The average C1–C2 and C2–C3 bond lengths in (IA), (IB) and (II) are 1.532 (1) and 1.534 (4) Å, respectively. These values are 0.01–0.02 Å greater than corresponding values in (VI), which are 1.522 (4) and 1.515 (2) Å, respectively, presumably reflecting the different substitutions at C2. All other corresponding C–C bonds in (IA), (IB) and (VI) have very similar lengths. Within the same three structures, the exocyclic C5–C6 bond appears to be the shortest C–C bond [1.516 (3) Å]. Similar inspections of the C1–O1 and C1–O5 bonds suggest a slight lengthening of the former and a slight shortening of the latter in N-acyl sugars (IA) and (IB) compared with the related bonds in the simple glycoside (VI).

In all structures in Table 1 bearing an N-acyl group, the average C2–N1 bond length is 1.453 (5) Å. This bond is ~ 0.03 Å longer than the average exocyclic (non-anomeric)

C–OH bond length in a pyranosyl ring, which is 1.427 (4) Å in the same data set.

Within the *N*-acyl exocyclic fragment, the C8–O8 bond averages 1.236 (5) Å, with no discernible difference between *N*-acetyl and *N*-formyl groups. The C8–C9 bond averages 1.502 (8) Å, which is 0.02–0.03 Å shorter than the endocyclic C–C bonds found in the pyranosyl rings.

The *N*-acyl side chains in (IA), (IB) and (II) contain the amide bond N1–C8 in a *trans* conformation, *i.e.* with C2–N1–C8–C9 torsion angles of ± 179.1 (2)° in both (IA) and (IB), and an average C2–N1–C8–O8 torsion angle of 0.77 (2)° in (IA), (IB) and (II). These torsion angles also demonstrate that the amide group is planar. In all three structures, the conformation about the C2–N1 bond is *anti*, *i.e.* atom N1H is roughly *anti* to H2, which is consistent with the behavior reported in solution (Zhu *et al.*, 2006). However, inspection of the C1–C2–N1–C8 and C3–C2–N1–C8 torsion angles in (IA), (IB) and (II) reveals a range of 91–108° in the former and –128 to –144° in the latter, indicating some flexibility about the C2–N1 bond in the solid state.

The C5–O5–C1 bond angles in (I)–(VI) appear to depend on anomeric configuration, with β -anomers yielding an average value of 112.0 (6)° and α -anomers an average of 114.7 (3)°. Within the full data set, the largest C–C–C bond angle within any given structure (β -anomers only) is C4– C5–C6, which averages 113.8 (11)°. The two bond angles that incorporate the carbonyl O atom, O8–C8–C9 and O8– C8–N1, are very similar in each structure and average 122.5 (14)° in the full data set. These results contrast with the remaining angle, N1–C8–C9, which is uniformly smaller than the others in any given structure and averages 116.0 (4)° in the full data set.

Within (IA), (IB) and (II), the endocyclic ring torsion angles vary from 41 to 72° (absolute values), indicating that aldopyranosyl rings containing *N*-acyl substitutents at C2 are distorted. It is noteworthy that the extreme angles within this group are observed in (II), where the C1-C2-C3-C4 and C1-O5-C5-C4 torsion angles are -41.09 (15) and 71.61 (13)°, respectively. Within (III)-(VI), these torsion angles range from 48 to 68° (absolute values). A more quan-





The atom-labeling scheme for compound (II). Displacement ellipsoids are drawn at the 50% probability level.



Figure 3 Hydrogen-bonding scheme for molecules (IA) and (IB), viewed along the *a* axis. Dashed lines (blue in the electronic version of the paper) represent hydrogen bonds.

titative treatment of these torsion angles is provided by the Cremer–Pople parameters calculated for (I)–(VI) (Table 2; Cremer & Pople, 1975). The most distorted ring within (I)–(VI) is that of (II), which yields a θ value of 16.59 (14)°. The least distorted ring is found in (Va), where $\theta = 0.9$ (3)°, and the ring is almost an ideal ${}^{4}C_{1}$ chair. The direction of distortion varies widely within this series of compounds. For (IA) and (II), $\varphi = 302.0$ (12) and 314.4 (5)°, respectively, suggesting distortion towards $B_{C2,C5}$. In contrast, φ for (IB) is 0 (2)°, or a ${}^{C3,O5}B$ distortion. These data show that not only can aldopyranosyl rings be substantially distorted when bearing an *N*-acyl functionality at C2, but the direction of distortion can also vary widely, with various boat ($B_{C2,C5}$, ${}^{C3,O5}B$ and $B_{C1,C4}$) and twist-boat (${}^{C1}TB_{C5}$, ${}^{C3}TB_{C1}$, ${}^{O5}TB_{C2}$ and ${}^{C5}TB_{C1}$) conformations represented.

The exocyclic hydroxymethyl conformation in each of (IA), (IB) and (II) is gt (C4 *anti* to O6), but the gg conformation (H5 *anti* to O6) is observed in each of (III), (IV*a*), (V*a*) and (V*b*). These data show that gg and gt conformations are favored in GlcNAc/NFm aldopyranosyl rings in the solid state, a behavior which is expected to mimic that in solution, based on related studies of the simpler Glcp anomers (Thibaudeau *et al.*, 2004).

The structure of (I) forms hydrogen-bonded pairs of (IA) and (IB) molecules of through hydroxy atom O6A to atom O6B. In turn, atom O6B forms a hydrogen bond to atom O6Aⁱⁱⁱ of the next pair related by translation along the *a* axis. In addition, the amide groups form hydrogen bonds, although this is to the same GlcNAc molecule related by translation along the *a* axis (all symmetry codes as in Table 3). Amide atom N1A of (IA) has a bifurcated hydrogen bond shared between the adjacent amide carbonyl atom O8Aⁱ and methoxy atom O1Aⁱ. Amide atom N1B of (IB) has a single hydrogen bond to amide atom O8Bⁱ. This association results in chains of pairs of (IA) and (IB) that run through the lattice parallel to the *a* axis. These chains are hydrogen bonded to other chains *via* hydroxy–hydroxy (O4B···O4Aⁱⁱ) or hydroxy–water inter-





Hydrogen-bonding scheme for (II), viewed along the a axis. Dashed lines (blue in the electronic version of the paper) represent hydrogen bonds.

actions. The water molecules are located in a hydrophilic channel lined with amide O atoms (O8B) and hydroxy groups, again parallel to the a axis. One motif that is apparent is a hydrogen-bonded ring formed by four sets of these hydrogenbonded chains. These rings are oriented around a hydrophobic channel formed by the methyl groups of the methoxy moieties and the acetamide methyl group (Fig. 3; see Table 3 for specific details). Overall, this structure forms a highly hydrogenbonded three-dimensional network of water and amidosaccharide molecules.

The structure of (II) also forms a three-dimensional network of hydrogen-bonded molecules. The network is formed from sheets of (II) that have hydrogen bonds through atom O4 to atom $O8^{iii}$ running parallel to the *b* axis (related by the 2_1 screw axis) and from atom O6 to atom $O4^{iv}$ along the *c* axis, and thus these sheets lie in the *bc* plane of the lattice. Propagation of these sheets into the third dimension is *via* hydrogen bonds from atom O3 to atom $O6^{ii}$ and from atom N1 to atom $O8^i$ of sheets in the next layer along the *a* axis (Fig. 4; all symmetry codes as in Table 4).

Experimental

Compounds (I) and (II) were prepared as described recently in the literature (Hu, Zhang *et al.*, 2010). Both compounds were crystallized from water at room temperature, giving long colorless needle-like crystals that were harvested for X-ray analysis.

Compound (I)

Crystal data $C_9H_{17}NO_6 H_2O$ $M_r = 253.25$ Orthorhombic, $P2_12_12_1$ a = 4.6886 (1) Å b = 14.4501 (3) Å c = 34.7880 (7) Å

$$\begin{split} V &= 2356.91 \ (8) \ \text{\AA}^3 \\ Z &= 8 \\ \text{Cu \mathcal{K}a$ radiation} \\ \mu &= 1.06 \ \text{mm}^{-1} \\ T &= 100 \ \text{K} \\ 0.25 \ \times \ 0.03 \ \times \ 0.02 \ \text{mm} \end{split}$$

Table 1

Comparison of geometric parameters in compounds (I)-(VI).

gg denotes a gauche-gauche conformation and gt gauche-trans.

Parameter	β -GlcNAc- OCH ₃ (IA)	β -GlcNAc- OCH ₃ (I <i>B</i>)	β -GlcNFm- OCH ₃ (II)	α-GlcNAc- OH (III)	β-GlcNAc- OH (IVa)	β-GlcNAc- OR (IVb)	α-GlcNAc- OH (Va)	β -GlcNAc- OR (Vb)	β -Glc- OCH ₃ (VI)
Bond lengths (Å)									
C1-C2	1.533 (3)	1.531 (3)	1.5318 (19)	1.534 (2)	1.522 (5)	1.522 (3)	1.526 (5)	1.515 (4)	1.522 (4)
C2-C3	1.530 (3)	1.534 (3)	1.5383 (19)	1.531 (3)	1.521 (4)	1.531 (4)	1.527 (5)	1.517 (5)	1.515 (2)
C3-C4	1.527 (3)	1.525 (3)	1.5302 (19)	1.521 (3)	1.531 (3)	1.516 (5)	1.520 (4)	1.516 (4)	1.529 (3)
C4-C5	1.526 (3)	1.523 (3)	1.5284 (18)	1.528 (3)	1.536 (4)	1.507 (5)	1.519 (5)	1.537 (4)	1.527 (4)
C5-C6	1.519 (3)	1.514 (3)	1.5137 (19)	1.514 (2)	1.501 (4)	1.499 (5)	1.512 (5)	1.517 (4)	1.516 (2)
C1-O1	1.389 (3)	1.387 (3)	1.3899 (17)	1.390 (3)	1.389 (4)	1.389 (4)	1.361 (5)	1.395 (4)	1.379 (2)
C1-O5	1.418 (3)	1.423 (3)	1.4198 (17)	1.434 (2)	1.427 (3)	1.429 (3)	1.418 (4)	1.414 (4)	1.433 (2)
C2-N1	1.456 (3)	1.455 (3)	1.4615 (18)	1.457 (2)	1.450 (3)	1.446 (3)	1.450 (4)	1.460 (4)	
C2-O2				. ,					1.426 (2)
C3-O3	1.430 (3)	1.424 (3)	1.4282 (16)	1.430 (2)	1.430 (4)	1.424 (4)	1.421 (5)	1.431 (4)	1.425 (4)
C4-O4/O1	1.424 (3)	1.425 (3)	1.4252 (17)	1.434 (2)	1.448 (3)	1.425 (3)	1.448 (3)	1.422 (4)	1.426 (2)
C5-O5	1.443 (3)	1.435 (3)	1.4349 (17)	1.448 (2)	1.429 (3)	1.436 (4)	1.438 (4)	1.427 (5)	1.440 (2)
C6-O6	1.430 (3)	1.430 (3)	1.4341 (17)	1.416 (3)	1.413 (4)	1.415 (5)	1.419 (5)	1.423 (5)	1.417 (4)
C7-O1	1.445 (3)	1.439 (3)	1.4391 (16)						1.430 (4)
C8-O8	1.237 (3)	1.235 (4)	1.2334 (18)	1.235 (2)	1.243 (4)	1.246 (4)	1.231 (4)	1.231 (4)	
C8-C9	1.508 (3)	1.510 (3)		1.508 (3)	1.497 (5)	1.496 (6)	1.490 (6)	1.506 (7)	
Bond angles (°)									
C1 - C2 - C3	111.22 (19)	109.71 (19)	112.42 (11)	110.11 (13)	111.5 (4)	110.9 (4)	110.0(2)	109.3 (3)	108.4(2)
$C_{2}-C_{3}-C_{4}$	111.5 (2)	111.9 (2)	112.95 (11)	110.88 (15)	109.6 (4)	112.1 (4)	109.7(3)	109.1(2)	110.82 (13)
$C_{3}-C_{4}-C_{5}$	107.7(2)	110.12 (19)	108.46 (10)	108.77 (15)	111.2 (4)	110.0 (4)	110.7 (3)	110.3 (3)	111.15 (15)
C4 - C5 - O5	106.79(19)	108.70 (19)	106.84 (10)	108.41 (13)	110.1 (4)	108.2(4)	109.3(2)	110.2(3)	108.5 (3)
$C_{5}-O_{5}-C_{1}$	111.9 (2)	111.8 (2)	111.09 (10)	114.97 (14)	112.6 (4)	112.1 (4)	114.5 (3)	112.9 (2)	111.55 (11)
05 - C1 - C2	110.80(19)	108.8(2)	111 39 (11)	109.24(15)	109.3(4)	1091(4)	109.6(3)	1100(2)	108 38 (18)
C4 - C5 - C6	113.9 (2)	112.3(2)	114 51 (11)	11477(16)	113 3 (4)	1155(4)	1143(3)	1131(3)	112 33 (16)
$C_{2}-N_{1}-C_{8}$	122.6(2)	121.8 (2)	12360(12)	122.17 (18)	122.9 (4)	124 9 (4)	1248(3)	122.0(3)	
N1 - C8 - C9	1161(2)	1164(2)	120100 (12)	116.01(18)	1163(4)	115.9 (5)	115 3 (3)	1158(4)	
08 - C8 - C9	121.3(2)	120.7(2)		120.93(16)	1222(4)	122.7(5)	120.7(3)	120.2(4)	
08-C8-N1	122.6 (2)	122.9 (2)	125.18 (13)	123.06 (16)	121.5 (4)	121.3 (4)	123.9 (3)	123.9 (3)	
Torsion angles (°)									
C1 $C2$ $C3$ $C4$	-47.0(2)	_50 1 (2)	-41.00 (15)	-546(2)	_52 1 (4)	_48.2 (4)	-544(4)	_567(2)	_54 2 (2)
C1 - C2 - C3 - C4	-47.9(3)	-30.1(3)	-41.09(13)	-34.0(2)	-32.1(4)	-46.2(4)	-34.4(4)	-30.7(3)	-34.3(3)
C1 = 03 = C3 = C4	55.1(2)	50.3(2)	/1.01 (15)	56.8(2)	50.4(4)	07.4 (4) 50.0 (4)	545(3)	54.2 (4)	51.7(2)
$C_2 = C_3 = C_4 = C_5$	55.1(5)	50.5(3)	49.70(13)	50.0 (2)	50.4 (4) 62.0 (4)	50.9 (4)	54.5(5)	54.2 (4) 62 1 (2)	51.7(5)
$C_2 = C_1 = O_3 = C_3$	-01.8(3)	-00.0(3)	-02.48(14)	-39.9 (2)	-02.9 (4)	-04.4(4)	-00.9 (3)	-02.1(3)	-07.9(3)
$C_2 = C_1 = O_1 = C_1/C_4$	173.3(2)	109.5(2)	170.03(11) 62.07(12)	58 4 (2)	54 8 (4)	131.7 (4) 58 6 (4)	55 7 (2)	54.7(4)	54.2 (2)
$C_{2} = C_{4} = C_{2} = 0.5$	-03.9(3)	-30.0(3)	-05.27(15)	-36.4(2)	-34.8(4)	-36.0(4)	-33.7(3)	-34.7(4)	-34.2(2)
$C_{2} = C_{2} = C_{1} = 0_{3}$	49.0 (3)	172.0(3)	43.79 (13)	35.6 (4) 177.51 (17)	37.0(4)	178 1 (4)	1745(2)	175.6(3)	171.0(2)
$C_{3} = C_{4} = C_{3} = C_{0}$	176.6 (2)	-175.9(2)	1/6.30(13)	-1/7.51(17)	-1/4.4(4)	-1/8.1(4)	-1/4.3(3)	-173.0(3)	-1/1.9(2)
03-03-00-00	04.5 (5)	03.2 (3)	00.48 (14)	-00.71 (18)	-00.0 (4)	38.0 (4)	-/4.0 (4)	-03.5 (3)	08.7 (S)
C1 - C2 - N1 - C8	g_i 108 2 (3)	g_l	g_l 91 34 (14)	<i>gg</i> 140 89 (17)	$gg_{100,5,(4)}$	g_l 1137 (4)	$\frac{gg}{138.7(3)}$	gg 100 5 (4)	gı
$C_{3}-C_{2}-N_{1}-C_{8}$	-1281(2)	-1372(2)	-144.20(13)	-968(2)	-1352(4)	-1225(4)	-989(4)	-1370(3)	
$C_2 - N_1 - C_8 - C_9$	179.1 (2)	-179.1(2)	117.27 (13)	169.86 (15)	-173.7(4)	178.4(5)	-179.6(4)	-173.9(4)	
	trans	trans		trans	trans	trans	trans	trans	
C2-N1-C8-O8	-1.2(4)	0.8 (4)	2.7 (2)	-9.7 (2)	5.2 (7)	-5.3(6)	-2.1(6)	2.9 (5)	

Data collection

Bruker APEX diffractometer
Absorption correction: multi-scan
(SADABS; Sheldrick, 2008)
$T_{\min} = 0.760, T_{\max} = 0.977$

Refinement

$$\begin{split} R[F^2 > 2\sigma(F^2)] &= 0.043 \\ wR(F^2) &= 0.111 \\ S &= 1.06 \\ 4311 \text{ reflections} \\ 323 \text{ parameters} \\ 6 \text{ restraints} \\ \text{H atoms treated by a mixture of} \\ \text{independent and constrained} \\ \text{refinement} \end{split}$$

21482 measured reflections 4311 independent reflections 3722 reflections with $I > 2\sigma(I)$ $R_{\text{int}} = 0.054$

 $\begin{array}{l} \Delta\rho_{\rm max}=0.20~{\rm e}~{\rm \AA}^{-3}\\ \Delta\rho_{\rm min}=-0.26~{\rm e}~{\rm \AA}^{-3}\\ {\rm Absolute~structure:~the~configura-tion~was~determined~based~on~the~known~handedness~of~the~chiral~C~atoms~within~the~structure\\ {\rm Flack~parameter:~-0.4~(2);~1726~Friedel~pairs} \end{array}$

Table 2

Cremer-Pople puckering parameters in compounds (I)-(VI).

Com- pound	θ (°)	$\varphi\left(^{\circ} ight)$	<i>Q</i> (Å)	q_2 (Å)	q_3 (Å)	Confor- mer†
(IA)	11.4 (3)	302.0 (12)	0.595 (3)	0.118 (3)	0.583 (3)	B_{C2} C5
(IB)	7.1 (2)	0(2)	0.585 (3)	0.078(2)	0.580(3)	C3,05B
(II)	16.59 (14)	314.4 (5)	0.5791 (14)	0.1654 (14)	0.5550 (14)	$B_{C2,C5}$
(III)	3.8 (2)	274 (3)	0.582 (2)	0.031 (2)	0.581 (2)	$^{C1}TB_{C5}$
(IVa)	4.8 (3)	19 (4)	0.568 (3)	0.044(3)	0.566 (3)	$^{C3}TB_{C1}$
(IVb)	8.7 (3)	338 (2)	0.577 (3)	0.089 (9)	0.570 (3)	$^{O5}TB_{C2}$
(Va)	0.9 (3)	55 (3)	0.572 (3)	0.006 (3)	0.572 (3)	$B_{C1,C4}$
(Vb)	2.3 (3)	97 (6)	0.580 (3)	0.029 (3)	0.580 (3)	$^{C5}TB_{C1}$
(VI)	6.94 (19)	38 (2)	0.597(2)	0.072(2)	0.593(2)	$^{C3}TB_{C1}$

† B denotes a boat conformation and TB a skew or twist-boat.

Compound (II)

Crystal data

 $\begin{array}{l} C_8 H_{15} \text{NO}_6 \\ M_r = 221.21 \\ \text{Monoclinic, } P2_1 \\ a = 4.5374 \ (5) \ \text{\AA} \\ b = 15.8837 \ (16) \ \text{\AA} \\ c = 6.8993 \ (7) \ \text{\AA} \\ \beta = 100.185 \ (4)^\circ \end{array}$

Data collection

Bruker APEX diffractometer Absorption correction: multi-scan (*SADABS*; Sheldrick, 2008) $T_{min} = 0.703, T_{max} = 0.940$

Refinement

$$\begin{split} R[F^2 > 2\sigma(F^2)] &= 0.027\\ wR(F^2) &= 0.070\\ S &= 1.06\\ 1744 \text{ reflections}\\ 137 \text{ parameters}\\ 1 \text{ restraint}\\ \text{H-atom parameters constrained}\\ \Delta\rho_{\text{max}} &= 0.16 \text{ e } \text{ Å}^{-3} \end{split}$$

Table 3

Hydrogen-bond geometry (Å, °) for (I).

$D - H \cdot \cdot \cdot A$	D-H	$H \cdots A$	$D \cdots A$	$D - H \cdots A$
$O3A - H3OA \cdots O1W^{i}$	0.84	1.85	2.672 (3)	166
$O4A - H4OA \cdot \cdot \cdot O2W$	0.84	2.01	2.806 (3)	158
$O6A - H6OA \cdots O6B$	0.84	2.01	2.837 (3)	170
$N1A - H1NA \cdots O8A^{i}$	0.88	2.50	3.132 (3)	129
$O3B - H3OB \cdot \cdot \cdot O2W^{ii}$	0.84	1.85	2.658 (3)	160
$O4B - H4OB \cdots O4A^{ii}$	0.84	2.07	2.898 (2)	167
$O6B - H6OB \cdot \cdot \cdot O6A^{iii}$	0.84	1.98	2.815 (3)	173
$N1B - H1NB \cdots O8B^{i}$	0.88	2.02	2.838 (3)	154
$O1W-H1WA\cdots O3A$	0.84(1)	2.02 (1)	2.843 (3)	167 (3)
$O1W-H1WB\cdots O3A^{iv}$	0.84 (1)	2.03 (1)	2.847 (2)	165 (3)
$O2W - H2WA \cdots O8A^{v}$	0.84 (1)	2.01 (1)	2.837 (2)	171 (3)
$O2W-H2WB\cdots O3B^{vi}$	0.83 (1)	1.90 (1)	2.728 (3)	174 (3)
C	1 (")	. 1		1

Symmetry codes: (i) x - 1, y, z; (ii) $-x, y + \frac{1}{2}, -z + \frac{1}{2}$; (iii) x + 1, y, z; (iv) $x + \frac{1}{2}, -y + \frac{3}{2}, -z$; (v) $x - \frac{1}{2}, -y + \frac{3}{2}, -z$; (vi) $-x + 1, y - \frac{1}{2}, -z + \frac{1}{2}$.

The absolute configurations of (I) and (II) were determined both from the known configuration of the starting materials and by comparison of the intensities of Friedel pairs of reflections. However, the Flack parameters were inconclusive [x = -0.4 (2) for (I) and 0.13 (14) for (II); Flack, 1983]. Further confirmation of the configurations was sought by the Hooft analysis, yielding a Hooft y parameter of -0.13 (12) and P2(true) and P3(true) values of 1.000 and 1.000 for (I), and a Hooft y parameter of 0.16 (4) and P2(true) and P3(true) values of 1.000 and 1.000 for (II) (Hooft *et al.*, 2008).

For both structures, the hydroxy, amide and, where applicable, water H atoms were all located from a difference Fourier map and initially included in those positions. The hydroxy and amide H atoms

$V = 489.40 (9) \text{ Å}^3$	
Z = 2	
Cu $K\alpha$ radiation	
$\mu = 1.11 \text{ mm}^{-1}$	
T = 100 K	
$0.34 \times 0.07 \times 0.06 \text{ mm}$	n

6422 measured reflections 1744 independent reflections 1744 reflections with $I > 2\sigma(I)$ $R_{int} = 0.021$

$$\begin{split} &\Delta \rho_{\rm min} = -0.21 \ e \ {\rm \AA}^{-3} \\ & {\rm Absolute \ structure: \ the \ configura-} \\ & {\rm tion \ was \ determined \ based \ on \ the} \\ & {\rm known \ handedness \ of \ the \ chiral \ C} \\ & {\rm atoms \ within \ the \ structure.} \\ & {\rm Flack \ parameter: \ 0.13 \ (14); \ 806} \\ & {\rm Friedel \ pairs} \end{split}$$

Table 4

H	ydrogen-	bond	geomet	try ((A, °)) for	(11))
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$D - H \cdots A$	D-H	$H \cdot \cdot \cdot A$	$D \cdots A$	$D - \mathbf{H} \cdots A$
$N1-H1\cdots O8^{i}$	0.88	2.29	3.0213 (16)	141
$N1 - H1 \cdots O1^i$	0.88	2.48	3.1621 (15)	134
$O3-H3\cdots O6^{ii}$	0.84	1.89	2.7271 (14)	173
$O4-H4\cdots O8^{iii}$	0.84	1.90	2.7438 (14)	177
$O6-H6\cdots O3^{iv}$	0.84	1.94	2.7749 (14)	171

Symmetry codes: (i) x - 1, y, z; (ii) x - 1, y, z - 1; (iii) $-x, y - \frac{1}{2}, -z$; (iv) x, y, z + 1.

were subsequently constrained to have reasonable geometric X-H bond distances and angles (N-H = 0.88 Å and O-H = 0.84 Å). Where applicable, mild restraints were applied to the water O-H bond distances [0.84 (1) Å]. All C-H bonds were constrained to distances of 0.98–1.00 Å. For all H atoms, $U_{\rm iso}({\rm H}) = 1.5U_{\rm eq}({\rm C})$ for methyl H atoms or $1.2U_{\rm eq}({\rm C})$ for all others.

Data collection: *APEX2* (Bruker–Nonius, 2009) for (I); *APEX2* (Bruker–Nonius, 2008) for (II). Cell refinement: *SAINT* (Bruker–Nonius, 2009) for (I); *SAINT* (Bruker–Nonius, 2008) for (II). Data reduction: *SAINT* (Bruker–Nonius, 2009) for (I); *SAINT* (Bruker–Nonius, 2008) for (II). For both compounds, program(s) used to solve structure: *SHELXS97* (Sheldrick, 2008); program(s) used to refine structure: *SHELXL97* (Sheldrick, 2008); molecular graphics: *XP* (Sheldrick, 2008) and *POV-Ray* (Cason, 2003); software used to prepare material for publication: *XCIF* (Sheldrick, 2008) and *publCIF* (Westrip, 2010).

Supplementary data for this paper are available from the IUCr electronic archives (Reference: FA3249). Services for accessing these data are described at the back of the journal.

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